

Microbiological study of otitis media

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Abstract

Otitis media is a destructive & persistent disease with irreversible sequelae. It is associated with bacterial or fungal aetiology.

Objectives: 1) To identify bacterial isolates causing ear infection and their antibiogram. 2) To identify fungal isolates 3) To screen all smears for Acid Fast Bacilli (AFB).

Materials and Methods: This study was conducted for 1 year. Pus was collected with sterile swab before commencement of antibiotics, from 98 patients of otitis media, attending ENT OPD. Gram's staining & Gabbet's staining (Modified ZN staining) was done for all the samples. Bacterial & fungal isolates were identified by standard methods. Antibiotic susceptibility testing of bacterial isolates was carried out by Kirby Bauer's disc diffusion method.

Results: Age of the patients studied, ranged from 2 to 68 years. Of the 98 patients, 48 were females and 50 were males. Bacterial isolates and their incidence were: Staphylococcus aureus(31%), CONS(6%), Pseudomonas species(20%), Klebsiella species(7%), Proteus species(5%). 4 fungi were isolated. 2 samples showed acid fast bacilli on Gabbet's staining.

Conclusions: The most common organism causing Otitis media in this study was Staphylococcus aureus (MSSA). Antibiotic treatment should be individualized depending on antibiotic sensitivity report, to reduce chronicity, complications & morbidity. It also showed in the study that, doing acid fast staining routinely could be of help as it helped to diagnose two cases of pulmonary tuberculosis.

Keywords: ENT- ear, nose, throat, OPD- out patient department, CONS- coagulase negative staphylococci, MSSA- methicillin sensitive staphylococcus aureus, AFB- acid fast bacilli

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management of otitis media and its complications and thus emergence of resistant bacterial strains can be prevented.⁵

Objectives

1. To know the bacterial etiology of CSOM
2. To know the fungal etiology of CSOM
3. To screen for acid fast bacilli in ZN smears.

Materials and Methods

Study was done for a period of one year from January to December 2014 in a District tertiary care hospital Mandya. Patients with history of ear discharge from 3 months were examined using aural speculum. Patients with central tympanic perforation who have not used any antibiotic in last 2 weeks were included in the study. Patients with history of antibiotic usage in 2 weeks were excluded. A total of 112 pus samples were received from the patients attending OPD of department of ENT. All samples were further processed in Department of Microbiology. 2 Sterile cotton swab was used to collect the ear discharge. Swab was first put for culture on Blood agar, MacConkey Agar plates and Saboraud Dextrose Agar slants. Other swabs were also used to make smears for Gram stain, ZN staining by Gabbet's method wet mount in 10% potassium hydroxide. All culture plates were incubated at 37^o C for 24 hours. All smears were examined under microscope within 30 minutes. All the cultures were identified by standard protocols.⁶ For fungal

Introduction

Chronic otitis media is one of the most common diseases, with highest prevalence in tropical countries like India¹. Chronic suppurative otitis media (CSOM) is defined as chronic inflammation of middle ear and mastoid cavity that may present with recurrent ear discharges for more than 3 months through a tympanic perforation.² It can occur in any age group but more common in childhood especially in lower socioeconomic group.³ It is a destructive & persistent disease with irreversible sequelae.²

Most common organisms associated are *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Proteus mirabilis*, *Klebsiella pneumoniae*, *Escherichia coli*, *Aspergillus sp* and *Candida albicans*.⁴ The rate of complications have become less common, since the introduction of antibiotics. However, due to irrational and increased use of wide-spectrum antibiotics, the resistance in the bacterial isolates has emerged. Therefore, microscopic examination of stained smears, culture and sensitivity will help in appropriate

identification Aspergillus Species and Candida albicans were used as controls.

Sensitivity of bacterial cultures was done by Kirby Bauer's disc diffusion method and Antibiotics were put according to CLSI guidelines.⁷ Antibiotics were obtained by HiMedia laboratories.

Results

Out of 112 samples, 90 showed single growth and 6 showed mixed growth. Of the 112 patients, 44 were females and 68 were males. Age of the patients studied, ranged from 2 to 68 years. *Staphylococcus aureus* (36.3%) was predominant organism isolated followed by *Pseudomonas aeruginosa* (31.4%). *Staphylococcus aureus* showed highest sensitivity to Vancomycin and Linezolid. *Pseudomonas aeruginosa* showed highest sensitivity to Amikacin and Imepenem. 2 samples showed acid fast bacilli on ZN staining. They were found to be positive for pulmonary tuberculosis retrospectively. Results are shown in tables.

Table 1: Percentagewise Distribution of growth pattern

| Type of Growth | Number of samples | Percentage |
|----------------|-------------------|------------|
| Single | 90 | 80.3 |
| Mixed | 06 | 5.4 |
| No growth | 16 | 14.3 |
| Total | 112 | 100 |

Table 2: Distribution of patients based on different age groups

| Age | Number (Percentage) |
|--------------|---------------------|
| < 10 years | 20 (17.8%) |
| 11-20 years | 40 (35.7%) |
| 21- 30 years | 24 (21.4%) |
| 31-40 years | 12 (10.7%) |
| 41-50 years | 10(8.9%) |
| 51-60 years | 04 (3.6%) |
| > 61 years | 02 (1.8%) |

Table 3: Sex distribution of patients

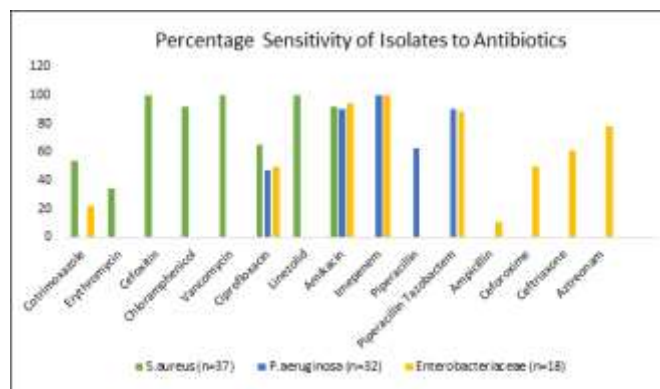
| Sex | Number | Percentage |
|--------|--------|------------|
| Male | 68 | 60.7% |
| Female | 44 | 39.3% |
| Total | 112 | 100 |

Table 4: Percentage distribution of organisms

| Microorganisms | Number | Percentage |
|----------------------------------|--------|------------|
| Staphylococcus aureus | 37 | 36.3% |
| Pseudomonas aeruginosa | 32 | 31.4% |
| Coagulase negative staphylococci | 3 | 2.9% |
| Steptococci | 3 | 2.9% |
| Klebsiella pneumoniae | 7 | 6.9% |
| Escherichiae coli | 6 | 5.9% |
| Proteus spp | 5 | 4.9% |
| Acinetobacter Sp | 2 | 1.9% |
| Diphtheroids | 3 | 2.9% |
| Aspergillus Niger | 3 | 2.9% |
| Candida Sp | 1 | 0.9% |

Table 5: Antibiotic Sensitivity pattern of *Pseudomonas aeruginosa* (32), *Staphylococcus aureus* (37) and Enterobacteriaceae family- *E.coli*, *Klebsiella Sp*, *Proteus Sp* (18)

| | Cot | Cip | E | LZ | Cx | Va | AK | C | CAZ | Pi | PiT | IPM | Amp | CTR | CXM | AZT |
|--|-----|-----|----|----|----|----|----|----|-----|----|-----|-----|-----|-----|-----|-----|
| S.aureus (n=37) | 20 | 24 | 34 | 37 | 37 | 37 | 34 | 34 | - | - | - | - | - | - | - | - |
| P.aeruginosa (n=32) | - | 15 | - | - | - | - | 29 | - | 16 | 20 | 29 | 32 | | | | |
| E.coli, Proteus, Enterobacteriaceae (n=18) | 4 | 9 | - | - | - | - | 17 | - | - | - | 16 | 18 | 2 | 11 | 9 | 14 |

**Fig. 1 : Percentage Sensitivity of Isolates to Antibiotics**

Discussion

India has been identified as one of the high prevalent countries for chronic SOM. One of the reason for this kind of prevalence could be that SOM is often considered as a part of normal.

Childhood phenomenon and people tend to tolerate the disease and live with its complications into adult life. Children constitute the most vulnerable group on account of their frequent exposure to upper respiratory tract infections. CSOM is a major cause of preventable hearing loss in developing countries. In children this may affect speech, psychological adaptability, cognitive development etc.⁸

Adults face the risk of social stigma and decreased avenues of employment.

In the present study most of the isolates were from patients in the age groups of 10-20 years (35.7%) followed by 20-30 years (21.4%) and indirectly signal acute exposure during the childhood. Similar age pattern has been reported by

Mansoor et al.⁹, and Poorey et al.¹⁰ In contrast, Loy *et al* showed the increased prevalence of CSOM in 30 - 40 years age in his study.¹¹

Predominant involvement of males (60.7%) over females (30.7%) in our study corroborates with data reported by other authors.^{12,13,14}

Managing CSOM cases on the basis of microbial causes, identification of aerobic, anaerobic and fungal isolates along with drug susceptibility testing is indispensable for taking appropriate clinical decisions. In our study, organisms were grown in 85.7% of the samples from CSOM patients which is similar to other study from India and other countries.^{15,16} Study found *S. aureus* as the most common (36.3%) isolate which correlates with study by Prakash, et al.² Other studies shows *P. aeruginosa* as the most common isolate in CSOM followed by *S. aureus*.^{17,18} Gulati et al. reported the most common isolate as *Klebsiella* spp.¹⁹ This shows that depending upon the climatic conditions, antibiotic usage & geographical factors the bacterial spectrum in CSOM varies with time as well as

from place to place. It is imperative to perform antibiotic sensitivity testing among isolates from CSOM because the antimicrobial resistance is not restricted to the hospital setting, rather manifesting at the community level also and CSOM is mainly the condition operating at the community level. Antibiotic sensitivity pattern of *P. aeruginosa* in our study revealed 100% susceptibility to Imepenem. Piperacillin-Tazobactam, showed 90% sensitivity among Gram negative isolates and Piperacillin was sensitive in only 62.5% of *P. aeruginosa*. Also 53 of the isolates were found resistant to Ciprofloxacin and 50% sensitivity to Ceftazidime. All the strains of *S. aureus* were 100% susceptible to Vancomycin, Teicoplanin, Cefoxitin and Linezolid. No strain was Methicillin Resistant *S. aureus* while 35-46% of *S.aureus* strains showed resistance to ciprofloxacin and cotrimoxazole. These findings are in comparison with previous studies^{17,18,20}. Amikacin and Erythromycin shows 92% sensitivity. Other Gram negative isolates i.e. *Escherichia coli*, *Klebsiella* spp, and *Acinetobacter* spp. were found 100% sensitive to Imipenem and 90% sensitive to Piperacillin-Tazobactam which is in comparison with other studies^{17,18}. Ciprofloxacin showed 50% resistance while Ampicillin showed 90% resistance.

Management of CSOM consists mainly of treating and eradicating infection and closure of tympanic membrane. Perforated tympanic membrane is a permanent threat of microbial attack of the middle ear and persistent infection is a cause of associated morbidity. Antimicrobials can be prescribed at the peripheral level but surgical intervention would require specialized facilities.

Conclusion

To choose the appropriate topical or systemic drug in individual cases antimicrobial susceptibility pattern of the isolates prevalent should guide the clinician. Changes in the sensitivity pattern of pathogenic organisms occurs as a result of irrational and widespread usage of antibiotics. Most common organisms were *S. aureus* and *P. aeruginosa* and these were less susceptible to drugs like Ciprofloxacin, Cotrimoxazole and Gentamicin which are used commonly. *Acinetobacter* spp. was found to be most resistant all antibiotics. For effective treatment and to prevent any complications, periodic evaluation of antimicrobial susceptibility pattern of microbial pathogens is mandatory. The role of ear toilet for better efficacy of topical antibiotics should be emphasized. By reducing the time lag between onset of symptoms and treatment, we can minimize complications of CSOM. Routine acid fast staining should be done in culture negative samples which can help to diagnose tuberculosis of systemic origin.

Consent: Not applicable.

Ethical Approval: Not applicable.

Competing Interests: Authors have declared that no competing interests exist.

References

1. World Health Organization (WHO), 2004. Chronic suppurative otitis media: burden of illness and management options. WHO Library Cataloguing-in-Publication Data, Geneva, Switzerland.
2. Prakash R, Juyal D, Negi V, Pal S, Adekhandi S, Sharma M, et al. Microbiology of Chronic Suppurative Otitis Media in a Tertiary Care Setup of Uttarakhand State, India. *N Am J Med Sci*. 2013 Apr;5(4):282-287.
3. Adhikari P, Sinha BK, Pokhrel NR, Kharel B, Aryal R, Ma J. Prevalence of Chronic Suppurative Otitis Media school children of Kathmandu district. *Journal of Institute of Medicine*. 2007;29(3):10-12.
4. Anwar-us-salam, Abid Sh, Abdulla EM. Suppurative Otitis in Karachi:An Audit of 510 Cases. *Pak J Otolaryn* 1997;13:66-69.
5. Agrawal A, Kumar D, Goyal A, Goyal S, Singh N, Khandelwal G. Microbiological profile and their antimicrobial sensitivity pattern in patients of otitis media with ear discharge. *Indian Journal of Otolaryn*. 2013;19(1):6-8.
6. Duiguil JP, Collee JG, Fraser Ag. Laboratory strategy in the diagnosis of infective syndromes. In Collee JG, marmion BP, Fraser ag, Simmon A. Mackie and Mcartny practical medical microbiology. 14th ed.London:1996.
7. Clinical and Laboratory Standard Institute. Performance Standards for Antimicrobial Susceptibility Testing. Pennsylvania, USA. 2013;1(1):M2A9.
8. Bluestone CD, Klein JO. Otitis Media in Infants and Children. Microbiology. 3rd ed. Philadelphia, PA: W. B. Saunders. 2001;79-1014.
9. Mansoor T, Musani MA, Khalid G, Kamal M. *Pseudomonas aeruginosa* in chronic suppurative otitis media: Sensitivity spectrum against various antibiotics in Karachi. *J Ayub Med Coll Abbottabad*. 2009;21:120-3.
10. Poorey VK, Lyer A. Study of bacterial flora in csom and its clinical significance. *Indian J Otolaryngol Head Neck Surg*. 2002;54:91-5.
11. Loy AH, Tan AL, Lu PK. Microbiology of chronic suppurative otitis media in Singapore. *Singapore Med J* 2002;43:296-9.
12. Kumar R, Srivastava P, Sharma M, Rishi S, Nirwan PS, Hemwani K, et al. Isolation and antimicrobial sensitivity profile of bacterial agents in chronic suppurative otitis media patients at Nims Hospital, Jaipur. *International Journal of Pharmacy and Biological Sciences*. 2013;3(4):265-269.
13. Malkappa KS, Kondapaneni S, Supam BR, Chakraverti KT. Study of bacterial isolates and their antibiotic susceptibility pattern in Chronic Suppurative Otitis Media. *Indian Journal of Otolaryn*. 2012;18(3):136-139.
14. Parveen SS, Rao JR. Aerobic bacteriology of Chronic Suppurative Otitis Media (CSOM) in a teaching hospital. *Journal of Microbiology and Biotechnology Research*. 2012;2(4):586-589.
15. Prayaga N, Moorthy S, Lingaiah J, Katari S, Nakirakanti A. Clinical Application of a Microbiological Study on Chronic Suppurative Otitis Media. *International Journal of Otolaryngology and Head & Neck Surgery*. 2013;2:290-294.

16. Aslam MA, Ahmed Z, Azim R. Microbiology and drug sensitivity patterns of chronic suppurative otitis media. *J Coll Physicians Surg Pak.* 2004;14:459-61.
17. Kumar H, Seth S. Bacterial and fungal study of 100 cases of chronic suppurative otitis media. *J Clin Diagn Res.* 2011;5:1224-7.
18. Shyamla R, Reddy SP. The study of bacteriological agents of chronic suppurative otitis media— aerobic culture and evaluation. *J Microbiol Biotechnol Res.* 2012;2:152-62.
19. Gulati SK. Investigative profile in patients of chronic suppurative otitis media. *Indian J Otol.* 1997;3:59-62.
20. Gaur RS, Mathew J, Varghese AM, Mathew GA, Chandrasekharan R, Anandan S. Microbiological pattern of ear swabs in chronically discharging ears in a Tertiary Care hospital in India. *Indian J Otol.* 2013;19:51-4.

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